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**Distinguishing between spring and autumn spawned herring
from daily increments, a preliminary report.**

By

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ABSTRACT

Otoliths from Norwegian Spring Spawning herring larvae, collected on a cruise in May 1985 and North Sea Autumn Spawned herring larvae, collected on a cruise in Desember 1987, were examined for daily increments. A greater distance from the otolith nucleus to the first observed increment was seen in Norwegian Spring Spawning herring larvae ($14.0 \pm 3.4 \mu\text{m}$) than in North Sea Autumn Spawned herring larvae ($5.0 \pm 1.6 \mu\text{m}$). In addition the results indicate that species from autumn and spring spawning stocks can be separated by a comparison of the increment widths in the otoliths. In a distance between 40 and 60 μm from the nucleus, the increments in the North Sea Autumn Spawned herring larvae had a width less than 1.5 μm , while the corresponding widths in spring spawners were greater than 2.0 μm .

INTRODUCTION

The complexity of herring stocks in the Skagerrak - Kattegat area has been described by Rosenberg and Palmen (1981). One of the problems in this area has been separation problems between outbursts of spring and autumn spawning herring. Several methods has been suggested and are in use, like combined analysis of meristic characters, total fish length, scale length and otolith length (Rosenberg and Palmen, 1981), the size of the first winter zone (Otterlind, 1987) and length frequency distribution

differences between the two subspecies. This paper looks at the position of the first daily increment, and the width of the increments in the otoliths as possible parameters to distinguish between larvae of the two different stocks. The regularity of ring deposition and the dependence of growth-rate of the increment width has been verified by the enclosure experiments reported by Moksness and Wespestad (in press). The growth rate of the larvae reflect the feeding condition and the temperature in the sea. A herring larvae that hatch in the spring experience good feeding conditions, while in the autumn the prey density is very much reduced. This is printed in the otoliths and therefore daily increment width is proposed as an useful parameter to distinguish between spring and autumn spawners.

MATERIAL AND METHODS

The materials were sampled on two different cruises. Larvae of Norwegian Spring Spawning herring were collected on a cruise with F/F "Eldjarn" on the Norwegian continental shelf north of 62°N, in May 1985. The larvae were sampled with a large midwater trawl and frozen onboard. The larvae used in this study (N = 11) were sampled west of the Sklinna bank (Fig. 1).

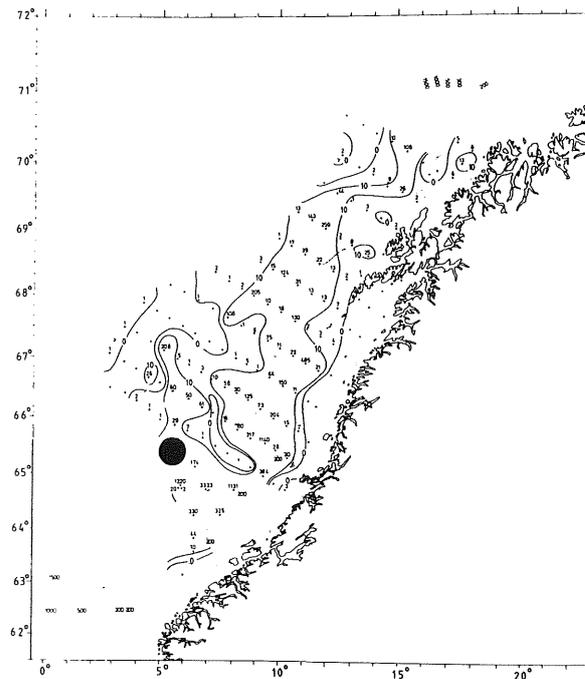


Figure 1. The herring larvae distribution, May 1985 (after Nedreaas 86).

● - Station 236 where the present larvae were sampled

Larvae from North Sea Autumn Spawners were collected on a cruise covering two transects between Denmark and Norway 9-13 Desember 1987 (Moksness and Johannessen 1988). All larvae were stored in 80% buffered ethanol and otoliths from a sample of these larvae were mounted on glass slides for later examination of daily increments.

The examination of the otoliths followed the same procedure as given in Andersen and Moksness (1988). An example of an otolith from larvae of spring spawning herring is given in fig. 2.

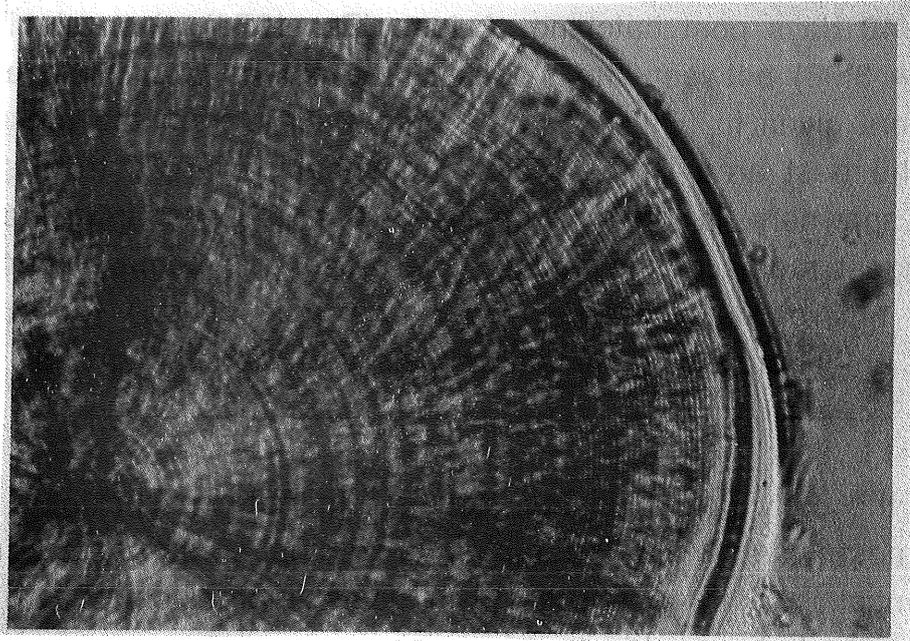


Fig. 2. Otolith of a 2 month old larvae of spring spawning herring.
200x magnification.

RESULTS

The distance observed from the nucleus to the first increment (Table 1) was $5 \pm 1.6 \mu\text{m}$ and $14 \pm 3.4 \mu\text{m}$ in the otoliths of larvae from autumn and spring spawning herring respectively. The reason for the difference in distance between the otolith nucleus and the first increment in spring and autumn spawners can be the reduced hatching-weight of larvae of autumn compared to larvae of spring spawners $100\text{-}120 \mu\text{g}$ compared to $180\text{-}200 \mu\text{g}$ (Blaxter and Hempel 1963).

Table 1. Mean distance between nucleus and first increment

Spring - spawners	SD	Autumn- spawners	SD
14.0 μm	3.4	5.0 μm	1.6

From figs. 3 and 4 it can be seen that the increment width in autumn spawners is persistent below 1.5 μm , with a mean about 1.0 μm in the whole period. In spring spawners, however, the increment width is above 2.0 μm from 40 days post hatching. A test, (Zar 1974) gives significant differences ($t= 3.06$) between the slopes of the curves of autumn and spring spawners given in figs. 3 and 4. It is evident from the present results that the best area in the otolith to separate larvae from spring and autumn spawners, is the area between 40 and 60 μm from the center, along the longest radius of the otolith. The reason for somewhat more variability in the results with spring spawners, may be due to that some of the larvae are approaching metamorphosis and then the growth is reduced.

Fig. 5. show the daily growth in the otoliths . In Autumn spawners this growth is almost linear, while the otolith growth in spring spawners is more exponential. All three figures (3-5) gives impression of large growth-differences between larvae of spring and autumn spawners.

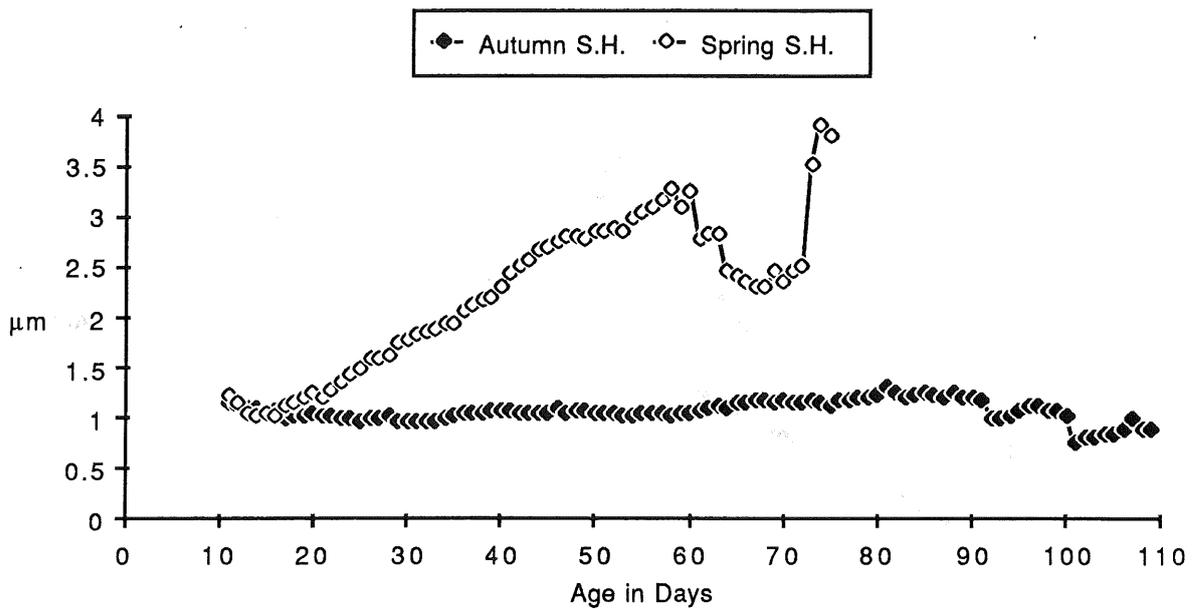


Fig. 3. Average increment widths in otoliths from North Sea Autumn Spawmed herring (Autumn S.H.) (N=31) and Norwegian Spring Spawmed herring (Spring S.H.) (N=11).

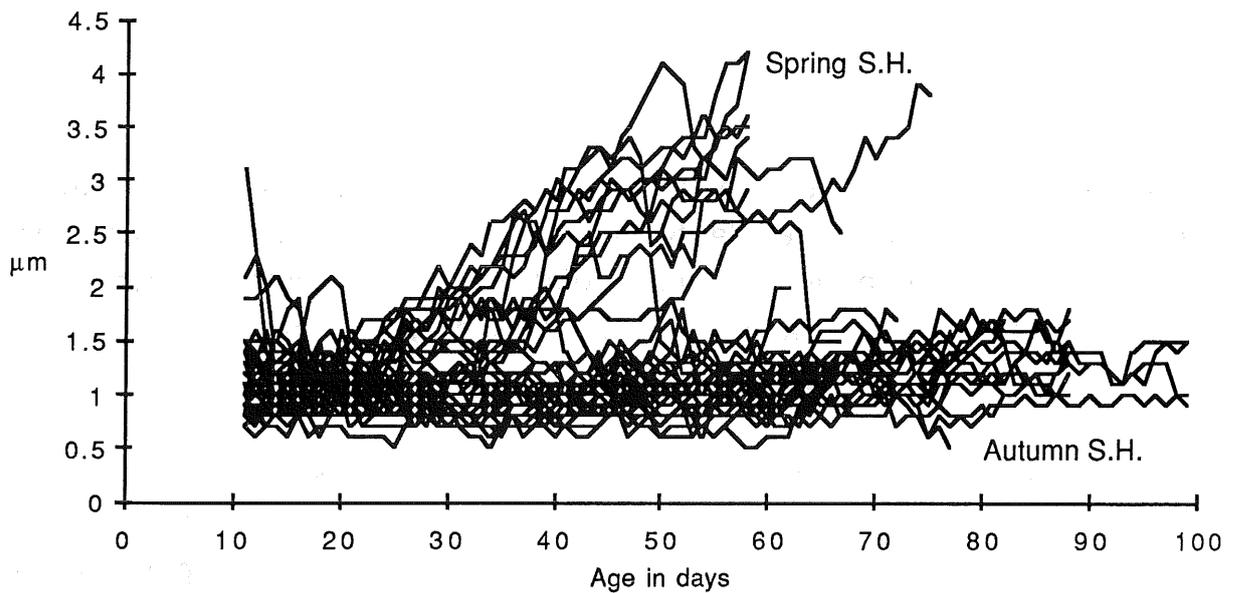


Fig. 4. Daily increment widths from each of the 31 North Sea Autumn Spawmed herring (Autumn S.H.) and 11 Norwegian Spring Spawmed herring (Spring S.H.).

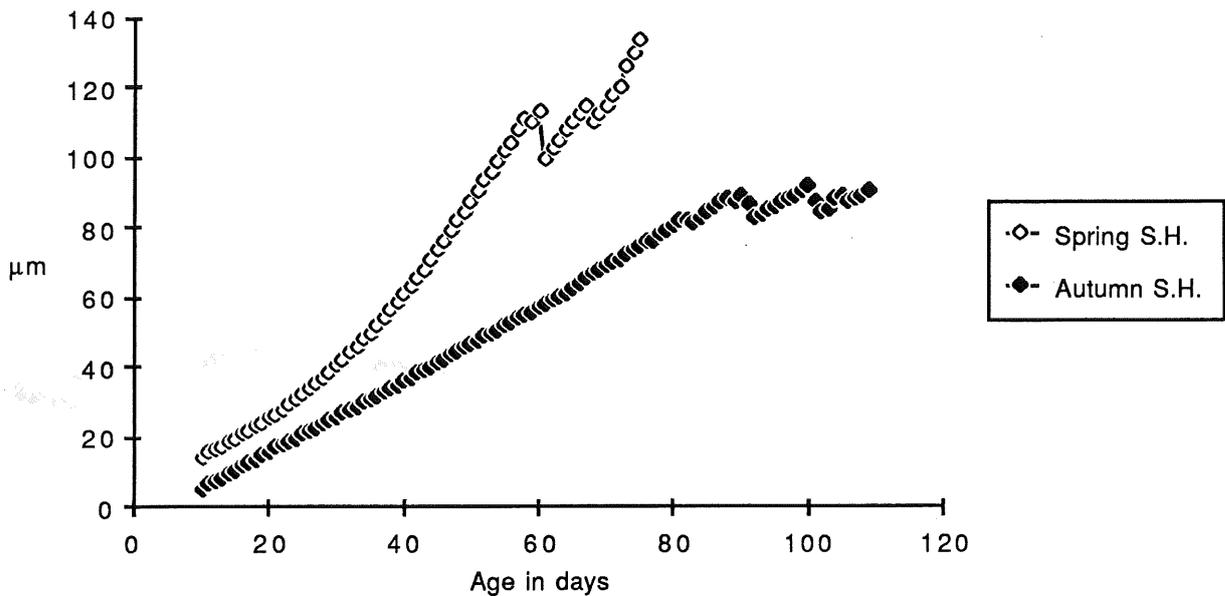


Fig. 5. Average distance from the otolith nucleus along the longest radius of the otolith as a function of age in the North Sea Autumn Spawmed herring (Autumn S.H.) (N=31) and Norwegian Spring Spawmed herring (Spring S.H.) (N=11).

DISCUSSION

Working with microstructure in otoliths, one is confronted with the crucial problem about regularity in ring deposition. Larvae exposed to cyclical stimuli experiences more distinct daily growth increment patterns than others (Pannella 1980). Such rythms can be vertical migration patterns, feeding regimes, light or temperature.

Growth dependant deposition have been shown to occure with herring larvae (Geffen 1983) kept in small volumes of water. Moksness et al.(1987) shows that herring larvae from Norwegian Spring spawmed in a laboratory experiment, fail to deposit growth increments untill their growth rate exceeds 0.15 mm per day, but later the deposition was regular. A recurring problem with lab. experiments may be that the larval growth are too low, or that the larvae lack a sort of calibrating mecanism. However, in experiments with herring larvae in mesocosms (Gjøsæter and Øiestad 1981, Moksness and Wespestad in press) ring deposition was found to be daily, and Moksness and Wespestad (in press) concludes that the increment width is significantly correlated with the

growth of the herring larvae. As the ring deposition rate is daily and increment widths depends on the growth rate, there is evident that the growth differences seen in situ, between larvae of spring and autumn spawners also are reflected in the otoliths.

The present paper deal with at least to separate stocks of herring , Norwegian Spring Spawning herring and North Sea Autumn Spawning herring, the latter one built up of several separate stock units spawning from August - Desember.

The growth rate of the larvae reflect the feeding condition and the temperature in the sea. A herring larvae that hatch in the spring is exposed to large environmental variations. The larvae experience a spring bloom with high densities of microzooplankton, and grow up with large cohorts of copepod nauplii and copepodites. The temperature is also improving through this period and everything is prepared for rapid growth and development.

The relative success of herring stocks in different ecosystem is dependant of the size and productivity of the system (Iles and Sinclair 1982). If the productivity of the system is too low, spring hatched herring larvae are not able to metamorphose through the metamorphose-window in june-august . The herring living in such a system will through selection be forced against late summer and autumn spawning. In the autumn the abundance of zooplankton, the temperature and light intensity are decreasing and Jones (1985) used daily increment analyses of otoliths to distinguish between herring larvae that had hatched early and late in the hatching period. After a short period of rapid growth, the growth rate is heavily reduced, and in the winter period the larvae enters a sort of lethargic condition.

All this is printed in the otoliths and therefore daily increment width is proposed as an useful parameter to distinguish between spring and autumn spawners in the present paper.

ACKNOWLEDGMENT

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