In vitro toxicity of hydrocarbon pollutants

AC Knag^{1*}, I Mayer², S Verhaegen², E Ropstad², S Mjøs³ and S Meier⁴

¹ Department of Biology, University of Bergen, Norway

² Department of Production Animal Clinical Sciences, Norwegian School of Veterinary Science

³ Department of Chemistry, University of Bergen, Norway

⁴ The Institute of Marine Research, Bergen, Norway

*Corresponding author: Phone: +47 970 46 302, E-mail: anne.knag@bio.uib.no

An investigation of the potential of polar oil related pollutants to disrupt or modulate steroidogenesis *in vitro*, using a human adrenocortical carcinoma cell line, the H295R assay.

INTRODUCTION Materials and methods Results

The human adrenocortical carcinoma cell line, H295R, expresses all the key enzymes for steroidogenesis, and thus is an ideal model to investigate effects of chemicals on steroid hormone production (Fig. 1).

To date, knowledge of how oil pollutants could impair fish reproduction, via their ability to disrupt steroidogenesis, the essential biochemical pathway responsible for the production of the sex steroid hormones, is not well studied. For this reason, we have investigated the potential of polar oil related pollutants to disrupt or modulate steroidogenesis *in vitro*, using a human adrenocortical carcinoma cell line, the H295R assay. In the first instance, this *in vitro* study focused on evaluating the deleterious effects of naphthenic acids (NA), alkyl

Cells were exposed to short chained AP (C2 and C3) with a similar composition as North Sea produced water (station 3 data, Harman et al., 2009), a commercial naphthenic acid mixture, and the polar fraction extracted from a produced water sample. The exact compositions of the exposure compounds were determined by gas chromatography. See Fig. 2 for AP composition of PW.

High = 10 mg/L (PW=4 mg/L)

Med = 1mg/L (PW=0,4mg/L)

Low = 0,1mg/L (PW=0,04mg/L)

After 48 hours exposure medium was collected and analyzed for hormone production (estradiol, testosterone and progesterone). Hormone concentrations were obtained by solid-phase radioimmunoassay kits. None of the exposure concentrations were cytotoxic according to Alamar Blue

ESTRADIOL

A significant increase in estradiol production was detected in the cells exposed to the highest dose of oil pollutants. See figure 3: Comparisons with medium collected from unexposed cells (MB) using the Tukey test, *= p<0.06, **=p<0,03



Figure 3. Estradiol production in cells exposed to hydrocarbon pollutants.

PROGESTERONE

TESTOSTERONE

A significant increase in progesterone production was detected in the cells exposed to the highest dose of oil pollutants. See figure 4: Comparisons with medium collected from unexposed cells (MB) using the Tukey test, ***= p<0.003

No significant alterations in testosterone produc-

tion in the cells exposed to oil pollutants (using

the Kruskall-Wallis test), see figure 5.



phenols (AP) and the polar fraction of produced water (PW) on steroid hormone production. The results of this *in vitro* study will indicate the potential of oil pollutants to disrupt endocrine function in wildlife, specifically the vitally important process of steroidogenesis.

assays.



Figure 2. Distribution in % of alkylphenols present in the extracted sample of PW. Total amount of AP in the extracted polar fraction of PW is 64%.

Figure 4. Progesterone production in cells exposed to hydrocarbon pollutants.



Figure 5. Testosterone production in cells exposed to hydrocarbon pollutants.



Naphthenic acids, C2 and C3 alkylphenols and the polar fraction of produced water induce the production of both estradiol and progesterone in exposed H295R cells. This indicates that polar hydrocarbons have the ability to modulate the steroidogenesis.





Figure 1. Steroidogenic pathway in H295R cells, adapted from Hecker & Giesy, 2008

Enzymes. **Hormones.** Corticosteroid pathways/products. Sex steroid pathways/products.

REFERENCES

HARMAN, C., THOMAS, K. V., TOLLEFSEN, K. E., MEIER, S., BØYUM, O. & GRUNG, M. (2009) Monitoring the freely dissolved concentrations of polycyclic aromatic hydrocarbons (PAH) and alkylphenols (AP) around a Norwegian oil platform by holistic passive sampling. Marine Pollution Bulletin, 58, 1671-1679.

HECKER, M. & GIESY, J. (2008) Novel trends in endocrine disruptor testing: the H295R Steroidogenesis Assay for identification of inducers and inhibitors of hormone production. Analytical and Bioanalytical Chemistry, 390, 287-291.

ACKNOWLEDGEMENTS

Thanks to Ellen Dahl at the Norwegian School of Veterinary Science for assistance with the RIAs. The Norwegian Research Council and the StatoilHydro travel fund is thanked for its financial support.

